



UNIVERSITI PUTRA MALAYSIA

**GENETIC DIFFERENTIATION OF MALAYSIAN OAKS BY
MICROSATELLITE MARKERS**

PUNG CHAI CHIN

FSAS 2001 24

**GENETIC DIFFERENTIATION OF MALAYSIAN OAKS BY
MICROSATELLITE MARKERS**

**BY
PUNG CHAI CHIN**

**Thesis Submitted in Fulfilment of the Requirement for the Degree of Master
Science in the Faculty of Science and Environmental Studies
Universiti Putra Malaysia**

January 2001



Abstract of thesis presented to the Senate of Univeristi Putra Malaysia in fulfilment of the requirement for the degree, Master of Science.

GENETIC DIFFERENTIATION OF MALAYSIAN OAKS BY MICROSATELLITE MARKERS

PUNG CHAI CHIN

January 2001

Chairman : Professor Dr. Tan Soon Guan

Faculty : Science and Environmental Studies

A combination of nuclear and chloroplast microsatellite DNA have been used to investigate the levels and pattern of variability in Malaysian oaks. This study focussed on *Quercus* sp., which is the largest and the most widely distributed genus in the family Fagaceae. In the nuclear microsatellite study, four sets of microsatellite primers developed for *Quercus* sp. (Dow *et al.*, 1995; Steinkellner *et al.*, 1997a; Isagi and Suhandono, 1997; Kampfer *et al.*, 1998) were tested. Based on the analysis, more than 35% of the 61 primers tested resulted in interpretable amplification products. Twenty microsatellite primers were used to estimate the genetic diversity among distributions of *Quercus* sp. These selected primers were also used in three other genera from the same family namely, *Lithocarpus*, *Castanopsis* and *Triganobalanus*.

The results showed that these microsatellite loci are conserved across different genera. Four primers, QpZAG9, QrZAG20, QrZAG31 and QrZAG108 gave interpretable PCR products for all the samples studied from the four genera. None of the microsatellite loci is monomorphic in all the species studied. The number of alleles per microsatellite locus varied from 2 to 20. On the average, 11.85 alleles per locus were

observed. The mean value of gene diversity ranged from 0.0141 at locus *QM50-3M* to 0.6494 at locus *QpZAG1/5* with a mean of 0.3162. The highest mean gene diversity (H_o) for all loci was 0.4290, which was observed in *Q. lineata* whereas the lowest H_o was found in *Castanopsis* sp. The genetic differentiation among the species was estimated as $F_{st} = 0.6705$. Three dendrograms based on Nei's genetic relationship (1978) clustered by the UPGMA method were constructed. The first dendrogram containing four different genera showed that *Lithocarpus* and *Castanopsis* are clustered in one group while *Triganobalanus* is clustered away from *Quercus*, *Lithocarpus* and *Castanopsis*. The second dendrogram showed that the main cluster is subdivided into two major subclusters. The Peninsula species was in one group whereas the Sarawak species formed another subcluster except for *Q. subsericea* from Banjaran Lumut. On the other hand, the third dendrogram which clustered individuals of all the species studied showed that all individuals from the same species is clustered together in the same group except for *Q. gemelliflora*. One unknown individual collected from the Kelabit Highlands is clustered together with *Q. sumatrana*. The estimates of genetic similarities based on microsatellite markers ranged from 0.0844 to 0.8590 among the different species.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Master Sains.

**PERBEZAAN GENETIK DALAM OAK MALAYSIA DIUJI DENGAN
PENANDA MIKROSATELIT.**

PUNG CHAI CHIN
January 2001

Pengerusi : Professor Dr. Tan Soon Guan

Fakulti : Sains and Pengajian Alam Sekitar

Satu gabungan kajian mikrosatelit nukleus dan kloroplas DNA digunakan untuk mengkaji paras dan corak variabiliti bagi oak Malaysia. Kajian ini memokus kepada *Quercus* sp. di mana ia merupakan genera yang terbesar dan terluas dalam famili Fagaceae. Dalam kajian mikrosatelit nukleus, sebanyak empat set primer yang direka untuk *Quercus* sp. (Dow *et al.*, 1995; ; Isagi and Suhandono, 1997; Steinkellner *et al.*, 1997a dan Kampfer *et al.*, 1998) diuji. Daripada analisis, lebih daripada 35% dari 61 primer yang diuji memberi produk amplifikasi yang boleh diinterpretasikan. Dua puluh primer digunakan bagi menjangka diversiti genetik untuk *Quercus* sp. Primer yang dipilih ini juga diuji dalam tiga genera dari famili yang sama iaitu *Lithocarpus*, *Castanopsis* dan *Triganobalanus*.

Keputusan menunjukkan lokus mikrosatelit adalah terpelihara antara genera yang berlainan. Empat primer iaitu QpZAG9, QrZAG20, QrZAG31 dan QrZAG108 memberi produk PCR yang boleh diinterpretasikan bagi semua sampel yang diuji berasal dari empat genera. Tiada lokus mikrosatelit yang monomorfik dalam semua sampel yang diuji. Nombor alel per penanda mikrosatelit adalah berlainan dari 2 hingga 20.

Secara puratanya, 11.85 alel diperhatikan. Purata diversiti gen pula berjulat dari 0.0141 pada lokus *QM50-3M* kepada 0.6494 pada lokus *QpZAG1/5* dengan purata 0.3162. Purata diversiti gen yang tertinggi (\bar{H}_o) dalam semua lokus ialah 0.4290, iaitu diperhatikan dalam *Q. lineata* manakala \bar{H}_o yang terendah diperhatikan pada *Castanopsis* sp. Perbezaan genetik antara spesis dijangka dengan $F_{st} = 0.6705$. Tiga dendrogram berdasarkan perkaitan genetik Nei's (1978) dikelompok berdasar cara UPGMA dikendalikan. Dendrogram yang pertama yang berdasarkan empat genera yang berlainan menunjukkan bahawa *Lithocarpus* dan *Castanopsis* dikelompokkan dalam satu kumpulan manakala *Triganobalanus* dikelompokkan jauh daripada *Quercus*, *Lithocarpus* dan *Castanopsis*. Dendrogram kedua menunjukkan kelompok utama di bahagikan kepada dua kelompok utama. Spesis Semenanjung dalam satu kumpulan manakala spesis Sarawak dalam satu kumpulan yang lain kecuali *Q. subsericea* dari Banjaran Lumut. Dalam pada itu, dendrogram yang ketiga yang mengelompokkan semua individu sampel dari semua spesis menunjukkan semua individu yang berasal dari spesis yang sama berada dalam satu kumpulan yang sama kecuali *Q. gemelliflora*. Satu individu yang tidak dikenali yang telah dikumpul dari Kelabit Highland adalah dikelompokkan bersama *Q. sumatrana*. Similariti genetik yang dijangka berdasarkan penanda mikrosatelit pada puratanya berjulat dari 0.0844 ke 0.8590 antara spesis yang berlainan.

AKNOWLEDGEMENTS

First and foremost, I would like to express my sincere appreciation and gratitude to my main supervisor Prof. Dr. Tan Soon Guan for his guidance, helpful comments, patience and suggestions. Without his help and supervision, this project would not have been possible.

Credit is also given to my supervisor Dr. Rusea Go. I learned many things from her especially in sample collections on several field trips and species identifications of my samples in this study. Without these, the project would also not have been possible.

I am also grateful to my supervisor, Associate Professor Datin Dr. Khatijah Mohd. Yusoff for giving me invaluable advice, in editing my thesis and also providing the facilities for the gel documentation and analysis in the virology laboratory.

A very special thank also goes to Prof. Dr. Soepadmo from FRIM, Kepong who helped me to identify my '*Quercus*' and also provided valuable comments for my project. Credit also goes to Dr. Lefort from the University of Crete who provided valuable comments in DNA isolation via emailing and Dr. Isagi who provided the primer sequences of the Japanese *Quercus*.

Besides, I would like to express my heartfelt thanks to Soon Choy and Chee Chin for their help in collecting the samples, organising the collected samples and also assisting me in laboratory work throughout the duration of this project.

I thank Mr Badrol from the Herbarium, Universiti Kebangsaan Malaysia for his technical assistance during my fieldwork. I also thank Julaihi, Jegung, Ah Tak, Michael and Lee from the Forestry Department of Sarawak for assisting me during the sample collection in Banjaran Lumut, Sarawak.

Special thanks also to Jacky Ho from UKM for sharing his valuable knowledge and providing me with valuable information on plant genetic research. Deepest gratitude also goes to my lab-mate Vijay who helped me a lot in data analysis. Permit me also to acknowledge the support, the helping, sharing and encouragement from all my lab-mates: Sahar, Boon Peng, Latif, Pau San, Bee Eng, Wan Ching, Lee Chien and Poh Kam. Our time of having lunch together is unforgettable.

Finally, I'm grateful beyond measure to my family and Siong for their unfailing love and support, I would also liked to thank everyone who had contributed throughout the duration of this study.

PUNG CHAI CHIN

2000/ Biology Department, Univeristi Putra Malaysia.

I certify that an Examination Committee met on 31st January 2001 to conduct the final examination of Pung Chai Chin on her Master of Science thesis entitled “ Genetic Differentiation of Malaysian Oaks by Microsatellite Markers” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that candidate be awarded the relevant degree. Members of Examination Committee are as follows:

Umi Kalsom, Assoc. Prof.
Lecturer,
Department of Biology
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Chairman)

Tan Soon Guan, Prof., J.S.M.
Lecturer,
Department of Biology
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Rusea Go, Ph.D.
Lecturer,
Department of Biology
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Datin Khatijah Mohd. Yusoff, Assoc. Prof.
Head,
Department of Biochemistry and Microbiology
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)



AINI IDERIS, Ph.D.
Professor
Dean of Graduate School,
Universiti Putra Malaysia

Date: 24 MAY 2001

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted fulfilment of the requirement for the degree of Master Science.



AINI IDERIS, Ph.D.

Professor

Dean of Graduate School,
Universiti Putra Malaysia

Date: **14 JUN 2011**

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which, have been duly acknowledged. I declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.



(Pung Chai Chin)

Date: 23/5/2001

TABLE OF CONTENTS

	PAGE
ABSTRACT	ii
ABSTAK	iv
ACKNOWLEDGEMENTS	vi
APPROVAL SHEETS	viii
DECLARATION FORM	x
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF PLATES	xviii
LIST OF ABBREVIATIONS	xx
CHAPTER	
1 INTRODUCTION	1.1
2 LITERATURE REVIEW	2.1
2.1 <i>Quercus</i> Linn.	2.1
2.2 <i>Lithocarpus</i> Blume.	2.13
2.3 <i>Castanopsis</i> (D. Don) Spach.	2.17
2.4 <i>Triganobalanus</i> Forman.	2.21
2.5 Genetic Studies on Oaks	2.24
2.6 Microsatellite DNA	2.30
2.7 Isolation of Microsatellite loci	2.34
2.8 Applications of Microsatellites	2.36
2.9 Chloroplast Microsatellite DNA	2.39
2.10 Hardy-Weinberg Equilibrium	2.40
2.11 Genetic Diversity	2.41
2.12 Polymerase Chain Reaction (PCR)	2.42
3 MATERIALS AND METHODS	3.1
3.1 Sample Collection	3.1
3.2 DNA Isolation	3.1
3.3 Primer Pairs	3.4
3.4 PCR Amplification	3.4
3.5 Characterisation of Microsatellite Loci	3.5
3.6 Electrophoresis of PCR Products	3.5
3.7 Voltage Applied	3.6
3.8 Data Analysis	3.6
3.8.1 Allele Frequencies	3.7
3.8.2 Testing for Hardy-Weinberg Equilibrium	3.7
3.8.3 Degree of Heterozygosity, <i>H</i>	3.8
3.8.4 Gene Diversity Index	3.9



	PAGE
3.8.5 Genetic Distances	3.10
3.8.5.1 F-statistics and Gene Flow	3.10
3.8.5.2 Nei's Genetic Distances (1978)	3.11
3.8.6 Test of Robustness	3.12
4 RESULTS	4.1
4.1 Sample Collection	4.1
4.2 Species Identification	4.5
4.3 DNA Isolation	4.16
4.4 Primer Screening and PCR optimisation	4.17
4.4.1 Primer Selection	4.17
4.4.2 Primer Banding Pattern	4.20
4.5 Allele Frequency	4.25
4.5.1 Single Species Description	4.25
4.5.2 Multiple Species Description	4.28
4.6 Hardy-Weinberg Equilibrium (HWE)	4.40
4.7 Degree of Heterozygosity, H	4.42
4.7.1 Single Species Description	4.42
4.7.2 Multiple Species Description	4.43
4.8 F-statistics, F	4.46
4.9 Gene Flow, N_m	4.47
4.10 Cluster Analysis	4.52
4.10.1 Genetic Distance and Dendrogram of Genera Level Clustering.	4.52
4.10.2 Genetic Distance and Dendrogram of Species Level Clustering.	4.54
4.10.3 Individual Clustering	4.55
4.11 Test of Robustness	4.60
4.12 Chloroplast Microsatellite Analysis	4.62
4.12.1 Primer Screening and PCR optimization	4.62
4.12.2 Gene Diversity Index	4.69
4.12.3 Genetic Differentiation	4.69
5 DISCUSSION	5.1
5.1 DNA Isolation	5.4
5.2 PCR Optimization	5.6
5.3 Trouble Shooting	5.10
5.3.1 PCR Amplification	5.10
5.3.2 Banding Scoring	5.13
5.4 Testing for Hardy-Weinberg Equilibrium	5.15
5.5 Heterozygosity, H	5.16

	PAGE
5.6 F-statistics, F and Gene Flow, N_m	5.17
5.7 Genetic Distances	5.18
5.8 Test of Robustness	5.20
5.9 Chloroplast Microsatellite	5.21
6 CONCLUSION	6.1
REFERENCES	R.1
APPENDIX A	A.1
APPENDIX B	B.1
APPENDIX C	C.1
BIODATA OF AUTHOR	D.1

LIST OF TABLES

TABLE		PAGE
2.1	Ecology and Distribution (by species) for Malaysian oaks.	2.11
3.1	Sample collection of Malaysian oaks.	3.2
4.1	Locations and dates of sample collections.	4.2
4.2	PCR conditions and allele character.	4.18
4.3	Multiple population description: Overall allele number and heterozygosity.	4.45
4.4	Summary of F- Statistics and gene flow for all loci.	4.48
4.5	Summary of F-Statistics and gene flow for three populations of <i>Q. subsericea</i> from Cape Rachado, Pasir Panjang and Banjaran Lumut.	4.49
4.6	Summary of F- Statistics and gene flow for two populations of <i>Q. gemelliflora</i> from Pasoh and Kelabit Highlands.	4.50
4.7	Summary of F- Statistics and gene flow for two populations of <i>Q. elmeri</i> from Fraser's Hill and Cameron highlands.	4.51
4.8	Nei's (1978) unbiased measure of genetic identity (above diagonal) and genetic distance (below diagonal) for 4 different genera.	4.53
4.9	Nei's (1978) Genetic identity (above diagonal) and genetic distance (below diagonal) for 19 populations from 15 species.	4.56
4.10	Chloroplast sequences and PCR conditions.	4.64
4.11	Chloroplast microsatellite primers developed for the tobacco chloroplast genome was used to amplify the genome in this study.	4.65
4.12	Summary of gene diversity statistics for all loci.	4.70
5.1	PCR inhibitors.	5.10
5.2	Solutions for PCR amplification problems.	5.11

LIST OF FIGURES

FIGURE		PAGE
2.1	Distribution of <i>Quercus</i> throughout the world.	2.4
2.2	Distribution of <i>Lithocarpus</i> throughout the world.	2.14
2.3	Distribution of <i>Castanopsis</i> sp. throughout the world.	2.18
2.4	Microsatellite DNA detected by amplification using PCR.	2.31
2.5	Strand-slippage Replication.	2.33
2.6	Schematic of the steps required to clone microsatellite loci in newly studied species	2.35
3.1	DNeasy Plant Mini Procedure (DNeasy plant handbook, 1999).	3.3
4.1	Location of sample collections in this study which included samples from Peninsula Malaysia and Sarawak.	4.3
4.2a	Leaf morphology of 3 species of <i>Quercus</i> .	4.7
4.2b	Leaf morphology of the <i>Castanopsis</i> sp. and <i>T. verticillata</i> .	4.8
4.2c	<i>Q. elmeri</i> from the population of Fraser's Hill and <i>Q. chrysotrica</i> from Cameron Highlands.	4.9
4.2d	<i>Q. elmeri</i> from the population of Cameron Highlands.	4.10
4.2e	<i>Q. gemelliflora</i> from population of Kelabit Highlands, Sarawak and <i>Q. lineata</i> from Cameron Highlands.	4.11
4.2f	Three different species namely <i>Q. kerangasensis</i> , <i>Q. gaharuensis</i> and <i>Q. valdinervosa</i> found in Kelabit Highlands.	4.12
4.2g	Two different species of <i>Lithocarpus</i> from Cameron Highlands.	4.13
4.2h	Three species of <i>Quercus</i> .	4.14
4.2i	Three populations of <i>Q. subsericea</i> from three different place namely Cape Rachado (Port Dickson), Pasir Panjang (Port Dickson) and Banjaran Lumut (Sarawak).	4.15

FIGURE		PAGE
4.3	Observed and effective numbers of alleles for each locus for all samples.	4.29
4.4	Overall allelic frequencies at locus <i>QM58TGT</i> .	4.30
4.5	Overall allelic frequencies at locus <i>QM69-2M1</i> .	4.30
4.6	Overall allelic frequencies at locus <i>QM67-3M1</i> .	4.31
4.7	Overall allelic frequencies at locus <i>QM50-3M</i> .	4.31
4.8	Overall allelic frequencies at locus <i>MSQ13</i> .	4.32
4.9	Overall allelic frequencies at locus <i>QpZAG1/5</i> .	4.32
4.10	Overall allelic frequencies at locus <i>QpZAG16</i> .	4.33
4.11	Overall allelic frequencies at locus <i>QpZAG9</i> .	4.33
4.12	Overall allelic frequencies at locus <i>QpZAG15</i> .	4.34
4.13	Overall allelic frequencies at locus <i>QpZAG36</i> .	4.34
4.14	Overall allelic frequencies at locus <i>QpZAG110</i> .	4.35
4.15	Overall allelic frequencies at locus <i>QrZAG11</i> .	4.35
4.16	Overall allelic frequencies at locus <i>QrZAG15</i> .	4.36
4.17	Overall allelic frequencies at locus <i>QrZAG20</i> .	4.36
4.18	Overall allelic frequencies at locus <i>QrZAG25</i> .	4.37
4.19	Overall allelic frequencies at locus <i>QrZAG30</i> .	4.37
4.20	Overall allelic frequencies at locus <i>QrZAG31</i> .	4.38
4.21	Overall allelic frequencies at locus <i>QrZAG65</i> .	4.38
4.22	Overall allelic frequencies at locus <i>QrZAG108</i> .	4.39
4.23	Overall allelic frequencies at locus <i>QrZAG112</i> .	4.39

FIGURE		PAGE
4.24	Genetic relationships of four different genera namely <i>Quercus</i> , <i>Lithocarpus</i> , <i>Castanopsis</i> and <i>Triganobalanus</i> based on Nei's (1978) genetic distance clustered by UPGMA.	4.53
4.25	Genetic relationships for 12 species of <i>Quercus</i> , a single group of <i>Lithocarpus</i> sp., <i>Castanopsis</i> sp. and <i>Triganobalanus verticillata</i> based on Nei's (1978) genetic distance clustered by UPGMA.	4.58
4.26	Individual clustering of all (152) individuals studied based on Nei's (1978) genetic distance clustered by UPGMA using PopGene.	4.59
4.27	Majority-rule consensus tree produced after 1000 bootstrapping.	4.61
4.28	Diversity Indices at seven loci for 4 species namely <i>Q. lineata</i> , <i>Q. subsericea</i> (Pasoh), <i>Lithocarpus</i> sp. and <i>Castanopsis</i> sp.	4.71

LIST OF PLATES

PLATE	PAGE
2.1 Acorns of <i>Quercus lineata</i> .	2.5
2.2 Acorn of <i>Lithocarpus</i> sp. found in Cameron Highlands.	2.15
2.3 <i>Castanopsis</i> sp found in Cameron Highlands.	2.20
2.4 Nuts of <i>Castanopsis</i> sp. are edible.	2.20
4.1a A wild <i>Quercus lineata</i> found beside the road to Blue Valley, Cameron Highlands.	4.4
4.1b <i>Lithocarpus</i> sp. found beside the road, Cameron highlands.	4.4
4.1c A single <i>Quercus elmeri</i> found on Fraser's Hill.	4.4
4.1d <i>Q. gemelliflora</i> from Pasoh 50 ha Plot forest reserve.	4.4
4.2 PCR products of <i>QrZAG30</i> showing monomorphism in <i>Q. argentata</i> with a molecular weight of 162bp.	4.21
4.3 PCR products of locus <i>QrZAG30</i> showing triple bands in <i>Q. subsericea</i> from Port Dickson population.	4.21
4.4 PCR products of locus <i>QpZAG9</i> for three species namely <i>Lithocarpus</i> sp. (1), <i>Castanopsis</i> sp. (2-9) and <i>T. verticillata</i> (10-14).	4.22
4.5 PCR products of locus <i>QrZAG20</i> in <i>Q. argentata</i> .	4.22
4.6 PCR products of locus <i>QrZAG31</i> in three species namely <i>Q. elmeri</i> from Cameron Highlands population (1-3), <i>Q. subsericea</i> from Banjaran Lumut population (4-15) and <i>Lithocarpus</i> sp. (16).	4.23
4.7 PCR products of <i>Q. elmeri</i> from Fraser's Hill population.	4.23
4.8 Monomorphic locus, <i>QrZAG65</i> , found in <i>Q. lineata</i> with a molecular weight of 218bp.	4.24
4.9 PCR products of locus <i>QrZAG112</i> of <i>Q. elmeri</i> from the Fraser's Hill population.	4.28

PLATE		PAGE
4.10	PCR products amplified by ccmp3, only one sample was a variant.	4.66
4.11	PCR products of ccmp5 in <i>Lithocarpus</i> sp.	4.66
4.12	PCR products of ccmp4 in <i>Q. subsericea</i> from Banjaran Lumut (1) and <i>Lithocarpus</i> sp. (2-14)	4.67
4.13	PCR products of ccmp6 in <i>Lithocarpus</i> sp.	4.67
4.14	PCR products of ccmp2.	4.68
4.15	PCR products of ccmp4.	4.68
5.1	Non-specific bands resulting from substrate excess conditions.	5.9
5.2	Weak PCR products	5.12

LIST OF ABBREVIATIONS

1X	One time
A	Adenosine
bp	Base pair
C	Cytosine
cpSSR	Chloroplast Simple Sequence Repeat
D	Genetic Distances
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleotide
EDTA	Ethylenediaminetetracetic acid
G	Guanine
nSSR	Nuclear Simple Sequence Repeat
PCR	Polymerase Chain Reaction
PHYLIP	Phylogeny Inference Package
RFLP	Restriction Fragment Length Polymorphism
Sp	Species
SSR	Simple Sequence Repeat
T	Thymine
TBE	Tris borate EDTA
UV	Ultra violet
VNTR	Variable Number of Tandem Repeat

CHAPTER 1

INTRODUCTION

The occurrence of oak tree in Malaysia is often surprising to most people. It is widely distributed throughout Peninsular Malaysia, Sabah and Sarawak. In terms of classification and taxonomy, Malaysian oaks were well documented by Soepadmo (1966, 1968 and 1972), Keng (1969), Corner (1972), Ridley (1967) and Soepadmo *et al.* (2000) for Sabah and Sarawak *Quercus*.

Oak is a common name for trees of the genus *Quercus* in the European and Asian region. Oak also refers to the genus *Lithocarpus* in the Malesia region. It had been reported that locally *Quercus* sp. and *Lithocarpus* sp. were known as *Castanopsis*, another genus from the Fagaceae family. On the other hand, some *Quercus* sp. was earlier identified as *Lithocarpus* sp. (Corner, 1972). In fact, according to Soepadmo, identification based on morphological evidences suggest they are actually two different genera.

Oak wood is less famous for timber products and is only used in medium to heavy construction. However, the beautiful wood ray of the species is well known as a source of timber for the furniture and flooring industries. Some of the species have been tried for use in the cultivation of mushroom in Borneo. It has proven useful for controlling erosion when planted on steep slopes in mountainous regions (Sunarno *et al.*, 1995).

Studies of oaks in Malaysia were limited to taxonomy. Earlier investigations of relationships among Malaysian oak accessions were based on morphological characters. *Quercus* sp. and *Lithocarpus* sp. which occurred throughout Malaysia are normally distinguished using leaf and acorn morphology characters either for species or genus level identification, but there is no truly diagnostic character that can assign an individual tree or population to one or the other species with certainty.

Furthermore, morphological evidence should be avoided since they may not be reliable measure of genetic difference because of the influence of the environment on gene expression. However, the analysis of plant DNA allows the direct assessment of variation in the genotype. Direct diagnostic markers allow the immediate detection of alterations without uncovering the effects of the alteration.

At present, development of molecular markers provides additional information on plants. DNA data also provide superior information, and with the advent of the PCR method, it may eventually become the predominant class of data. Recently, microsatellite DNA has been proven to be very useful for the purpose of studying genetic diversity in forest tree species (Lefort *et. al.* 1999a, Ujino *et. al.* 1998, Dow and Ashley 1998). Microsatellites are stretches of DNA consisting of di-, tri-, or tetra-nucleotide repeats such as (AT)_n or (GT)_n that frequently extend up to 100 times. Polymorphisms in microsatellites result from differences in the number of these repeat units. They are highly variable and co-dominantly inherited and can be used in genetic diversity, ecological and evolutionary studies.

Microsatellite primers are usually confined to a single taxon from which the primers were developed. However, some examples have been described in which microsatellite loci are conserved in other closely related species (Ujino *et al.*, 1998, Echt *et al.*, 1999 & Lefort *et al.*, 1999), which have allowed the analysis of genetic diversity to be carried out in these related species. Steinkellner *et al.* (1997a) have described the conservation of microsatellite loci between *Quercus* species. Their results showed that microsatellites isolated from *Quercus petraea* are conserved in other *Quercus* species and even in other species within Fagaceae.

With the establishment of these nuclear microsatellite primers (Steinkellner *et al.*, 1997a; Isagi and Suhandono, 1997; Kampfer *et al.*, 1998 and Dow *et al.*, 1995) for *Quercus* sp., this study was focused on diversity and genetic differentiation within the *Quercus* sp. that occurred throughout Malaysia. The selected primers were also tested in other three genera namely *Lithocarpus* sp., *Castanopsis* sp. and *Triganobalanus* sp. which are from the same family, Fagaceae.

Microsatellites are not limited to the nuclear genome. They are also found in the chloroplast genome. The developers of chloroplast microsatellite (cpSSR) primers in dicotyledonous angiosperms (Weising and Gardner, 1999) suggested that these universal primers may serve as general tools to study chloroplast variation in angiosperms. Thus, this study also tested the capability of cpSSR primers for amplifying the Malaysian oak genome. The extend of cross-species amplification appears to be correlated with taxonomic distance. Therefore, microsatellites are able be used for phylogenetic studies through the construction of informative dendrograms based on the allelic frequencies of microsatellite loci.

The objectives of this study were:

1. To develop microsatellite markers that could distinguish diagnostically between species Malaysian oaks.
2. To develop methodologies for typing microsatellites as genetic markers in Malaysian oaks.
3. To examine whether primer pairs designed to amplify microsatellite loci in other oak species could be used to amplify marker loci in Malaysian oaks.
4. To examine whether primer pairs designed to amplify chloroplast microsatellite loci in angiosperms could be used to amplify marker loci in Malaysian oaks.
5. To determine genetic relationships among *Quercus* sp., *Lithocarpus* sp., *Castanopsis* sp. and *Trigonobalanus* by constructing a dendrogram.
6. To clarify the exact identity of the taxa of Malaysian oaks by determining their genetic distances and through constructing a dendrogram.
7. To investigate the population structure of each species by estimating the genetic diversity at the microsatellite loci.
8. To examine whether microsatellite markers are suitable for taxonomic studies by determining the relationships between the species studied.